

## Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (*Oryza sativa*) FARO44 Under Water Deficit Conditions

Lawan Gana Ali,<sup>1</sup> Abubakar Haruna<sup>1</sup>, Mustapha Abdulkarim<sup>1</sup>

<sup>1</sup>Department of Science Lab. Tech., Mai Idris Aloomo Polytechnic Geidam, Yobe State, Nigeria.

Corresponding author: [lawanganaali@gmail.com](mailto:lawanganaali@gmail.com) +2348086038476

### Abstract

Application of potassium nitrate, silicon dioxide and salicylic acid to seed are approaches that impart stress tolerance to germinating seeds and seedlings under stressful conditions. This study evaluated the effects of synergistic osmopriming and hormonal priming with 2.5% KNO<sub>3</sub>+3.5% SiO<sub>2</sub>, 3.5% SiO<sub>2</sub>+1 mM SA and 2.5% KNO<sub>3</sub>+1 mM SA on seed germination, seedling growth, antioxidant enzymes, protein, proline and malondialdehyde contents in 3-weeks old FARO44 rice seedlings under drought conditions induced by polyethylene glycol (PEG6000). Germination experiments were laid in a completely randomised design (CRD) with each treatment replicated five times. The results revealed that synergistic priming significantly seedling growth, antioxidant enzymes catalase, ascorbate peroxidase and superoxide dismutase activities in rice seedlings by 2 folds compared to unprimed seedlings under drought. Synergistic effect of priming methods improved protein and proline contents in rice seedlings while decreased the content of malondialdehyde. Osmopriming and hormonal priming with 2.5% KNO<sub>3</sub>+3.5% SiO<sub>2</sub>, 3.5% SiO<sub>2</sub>+1 mM SA as well as 2.5% KNO<sub>3</sub>+1 mM SA showed different responses, however, they were found to be efficient in increasing germination, growth of seedling, antioxidant enzymes' activities as well as biochemical characteristics in rice seedlings grown under water deficit conditions. Enhanced activities of antioxidants, proline content as well as low malondialdehyde content in rice seedlings correlate with improved drought tolerance. Therefore, pre-soaking of FARO44 in 2.5% KNO<sub>3</sub>+3.5% SiO<sub>2</sub>, 3.5% SiO<sub>2</sub>+1 mM SA or 2.5% KNO<sub>3</sub>+1 mM SA is recommended for fast germination, seedling growth and drought tolerance in arid regions.

**Keywords:** *Germination; water deficit; antioxidants; biochemical attributes; proline*

## Introduction

Rice is an important crop consumed by many societies across the world. It serves as an essential daily food for more than half of the world's population (Dien *et al.*, 2019). Considering the significance of rice in both developed and developing nations, there is the need for its adequate supply, poverty reduction as well as improvement of financial status of people and nations (Zheng *et al.*, 2016).

Environmental constraints such as drought, salinity and extreme temperatures affect germination and seedling growth, accounting for 50% yield loss and as such impede the global rice production (Dien *et al.*, 2019; Pandey & Shukla, 2015; Quan *et al.*, 2016). Water is a key resource for successful crop production and is a highly limited resource. Drought causes colossal loss to crop production worldwide and is therefore a severe threat to sustainable agriculture. Globally, 23 million hectares of rice produced using rainfall as source of water are affected by water deficit situation (Pandey & Shukla, 2015). Moreover, changing climate that affect the world caused by the anthropogenic release of greenhouse gases is aggravating the water deficit conditions upon crop production (Intergovernmental Panel on Climate Change, 2007). Providing food to teeming population under depleting water resources needs crop varieties which can adapt and thrive under dry environments (Pandey & Shukla, 2015).

Poor germination as well as establishment of seedlings in rice under broadcasting systems of rice production caused by water

deficit results in enormous yield loss to rice growers (Liu *et al.*, 2016). Water shortage at early stage of crop production is responsible for delayed germination as well as irregular establishment of seedling (Kaya *et al.*, 2006). Shortage of water coupled with other environmental stresses are responsible for rapid production of reactive oxygen species (ROS) that ruin RNA, lipids, DNA, carbohydrates and proteins in crops (Gill & Tuteja, 2010). Reactive oxygen species impairment known as oxidative stress is a critical harm for plants facing abiotic stresses such as water shortage (Price *et al.*, 1989). While responding to stresses, plants deploy antioxidant systems consisting of catalase, glutathione, superoxide dismutase, peroxidase, superoxide dismutase and ascorbic acid to convert the toxic ROS to harmless substances (Gill & Tuteja, 2010; Hussain *et al.*, 2016).

Numerous approaches were employed aimed at induction of pressure tolerance in different crops (Hussain *et al.*, 2016). Priming of seed is a practical, easy, efficient and affordable method of inducing quicker as well as even seedling development. The technique also heightened vigor of seedling and yield of agricultural facing pressures of water shortage and salt stress (Jisha *et al.*, 2013; Paparella *et al.*, 2015). It is a controlled soaking of seeds in solutions of salts, water, and hormones or exposing to magnetic conditions, heat aimed at stimulating crucial processes of germination before the radicle emerge and sowing the seeds (Jisha *et al.*, 2013; Thomas *et al.*, 2013). Rapid and quality germination of soaked seeds are due to

*Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (Oryza sativa) FARO44 Under Water Deficit Conditions*

stimulation of enzymes, reduction of imbibition period, accelerated repair and production of germination metabolites as well as osmotic modification (Ashraf & Foolad, 2005; Farooq et al., 2006; Hussain et al., 2015; Hussain et al., 2016). The common approaches of rice priming used to deal with many stresses include osmopriming, hormonal priming, hydropriming, nutrient priming, redox priming, chemical priming as well as magnetopriming (Jisha et al., 2013; Paparella et al., 2015).

Northern Nigeria grow rice commonly using rainfall for germinating seeds, this system faces dry spell problems. Drought gravely upsets rice crop since rice have small roots, a slight water deficit cause hasty stomatal closing resulting in cessation of photosynthesis, change in metabolism of carbohydrate, osmolyte accumulation and other organic acids (Ji et al., 2012; Kamoshita et al., 2004). Thus, drought stress is an impediment for rice production, it is more unsettling in areas that depend on rainfall. Thus, this paper studied the synergistic or combined potentials of seed priming using potassium nitrate, silicon dioxide and salicylic acid priming on seedling growth, antioxidant enzymes, and

biochemical attributes in FARO44 rice under drought stress.

## Results

### Synergistic effects of osmopriming and hormonal priming on FARO44 rice seedling under drought condition

Osmopriming and hormonal priming had significant ( $P \leq 0.05$ ) effects on seedling length, seedling biomass and root length in FARO44 rice under drought stress (Table 1). Synergistic priming using 2.5%  $KNO_3$ +3.5%  $SiO_2$ , 3.5%  $SiO_2$ +1 mM SA or 2.5%  $KNO_3$ +1 mM SA significantly improved seedling length in FARO44 rice under drought stress compared with control. However, priming with 2.5%  $KNO_3$ +3.5%  $SiO_2$  produced significantly higher seedling under drought stress compared to other priming treatments. Similarly, priming using 2.5%  $KNO_3$ +3.5%  $SiO_2$ , 3.5%  $SiO_2$ +1 mM SA or 2.5%  $KNO_3$ +1 mM SA significantly enhanced rice seedling biomass and root length compared with control under drought stress. Priming with 2.5%  $KNO_3$ +3.5%  $SiO_2$  significantly improved root length of rice subjected to drought in comparison to other priming treatments evaluated in this study.

**Table 1: Seedling growth in FARO44 rice primed with combinations of  $KNO_3$ ,  $SiO_2$  and SA under PEG-induced drought stress**

Priming Treatments	SL (cm)	SB (Mg)	RL (cm)
Control	2.00 ± 0.03 <sup>d</sup>	51.91 ± 1.04 <sup>b</sup>	0.88 ± 0.04 <sup>d</sup>
2.5% $KNO_3$ +3.5% $SiO_2$	5.76 ± 0.09 <sup>a</sup>	52.48 ± 0.95 <sup>ab</sup>	4.79 ± 0.10 <sup>a</sup>
3.5% $SiO_2$ +1 mM SA	4.53 ± 0.10 <sup>b</sup>	52.43 ± 0.46 <sup>ab</sup>	3.61 ± 0.10 <sup>b</sup>
2.5% $KNO_3$ +1 mM SA	3.12 ± 0.02 <sup>c</sup>	54.34 ± 0.41 <sup>a</sup>	2.19 ± 0.03 <sup>c</sup>
Levels of Significance	0.000	0.000	0.000

Means and standard error (SE) in same column having similar superscripts are statistically the same as shown by Duncan Multiple Range Test at  $P \leq 0.05$ ; SL stands for Seedling length; SB = Seedling biomass; RL = root length.

*Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (Oryza sativa) FARO44 Under Water Deficit Conditions*

### Synergistic potentials of osmopriming and hormonal priming on superoxide dismutase, catalase and ascorbate peroxidase activities in FARO44 rice seedlings under drought stress

Osmo-hormonal priming had significant ( $P \leq 0.05$ ) effects on superoxide dismutase and catalase activity in FARO44 seedlings subjected to drought stress (Table 2). However, osmo-hormonal priming showed no substantial effect on ascorbate peroxidase activities in rice seedlings under drought stress (Table 3). Synergistic priming with 2.5%  $\text{KNO}_3$ +3.5%  $\text{SiO}_2$ , 3.5%  $\text{SiO}_2$ +1 mM SA and 2.5%  $\text{KNO}_3$ +1 mM SA significantly improved superoxide

dismutase and catalase activity of rice seedlings under drought stress compared to control. However, osmo-hormonal priming treatments had no effects in improving activities of ascorbate peroxidase in rice seedlings under drought compared with control. However, osmo-hormonal priming with 3.5%  $\text{SiO}_2$ +1 mM SA produced significantly higher catalase activity in rice seedlings under drought stress in comparison to other priming treatments.

**Table 2: Superoxide dismutase, catalase and ascorbate peroxidase activities of FARO44 rice seedlings primed with combinations of  $\text{KNO}_3$ ,  $\text{SiO}_2$  and SA under PEG-induced drought**

Priming Treatments	SOD (U/mgFW)	CAT (U/mg <sup>-1</sup> min <sup>-1</sup> FW)	APX (U/mg <sup>-1</sup> min <sup>-1</sup> FW)
Control	0.06 ± 0.02 <sup>c</sup>	0.00 ± 0.00 <sup>c</sup>	0.08 ± 0.01 <sup>a</sup>
2.5% $\text{KNO}_3$ +3.5% $\text{SiO}_2$	0.16 ± 0.03 <sup>bc</sup>	0.04 ± 0.01 <sup>b</sup>	0.14 ± 0.02 <sup>a</sup>
3.5% $\text{SiO}_2$ +1 mM SA	0.46 ± 0.02 <sup>a</sup>	0.09 ± 0.01 <sup>a</sup>	0.43 ± 0.22 <sup>a</sup>
2.5% $\text{KNO}_3$ +1 mM SA	0.25 ± 0.06 <sup>b</sup>	0.01 ± 0.00 <sup>bc</sup>	0.23 ± 0.06 <sup>a</sup>
Levels of Significance	0.000	0.000	0.197

Means and standard error (SE) in same column having similar superscripts are statistically the same as shown by Duncan Multiple Range Test at  $P \leq 0.05$ ; SL stands for Seedling length; SB = Seedling biomass; RL = root length

### Synergistic effects of osmopriming and hormonal priming on protein, proline and malondialdehyde contents in FARO44 rice seedlings under drought

Synergistic osmo-hormonal priming had significant ( $P \leq 0.05$ ) effects on protein, proline and malondialdehyde content of FARO44 rice seedlings as shown in Table 3. Osmo-hormonal priming with 2.5%  $\text{KNO}_3$ +3.5%  $\text{SiO}_2$  and 2.5%  $\text{KNO}_3$ +1 mM SA produced significantly higher content of protein in rice seedlings subjected to

drought stress in comparison with 3.5%  $\text{SiO}_2$ +1 mM SA.

Synergistic osmo-hormonal priming with 2.5%  $\text{KNO}_3$ +3.5%  $\text{SiO}_2$ , 3.5%  $\text{SiO}_2$ +1 mM SA and 2.5%  $\text{KNO}_3$ +1 mM SA significantly improved proline content and reduced the content of malondialdehyde in rice seedlings subjected to drought stress in comparison with control. However, osmo-hormonal priming with 2.5%  $\text{KNO}_3$ +3.5%  $\text{SiO}_2$  and 2.5%  $\text{KNO}_3$ +1 mM SA significantly enhanced proline content in

*Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (Oryza sativa) FARO44 Under Water Deficit Conditions*

rice seedlings compared with 3.5% SiO<sub>2</sub>+1 mM SA. Equally, osmo-hormonal priming manifestly reduced content of MDA in rice seedlings compared with control.

**Table 3: Protein, proline and malondialdehyde contents in FARO44 rice seedlings primed with combinations of KNO<sub>3</sub>, SiO<sub>2</sub> and SA under PEG-induced drought**

Priming Treatments	Protein (mg/ml)	Proline (mg/ml)	MDA (mM/mg <sup>-1</sup> FW)
Control	0.35 ± 0.03 <sup>c</sup>	3.22 ± 1.27 <sup>c</sup>	0.070 ± 0.00 <sup>a</sup>
2.5% KNO <sub>3</sub> +3.5% SiO <sub>2</sub>	0.73 ± 0.01 <sup>a</sup>	15.51 ± 1.63 <sup>a</sup>	0.001 ± 0.00 <sup>b</sup>
3.5% SiO <sub>2</sub> +1 mM SA	0.63 ± 0.03 <sup>b</sup>	9.79 ± 2.26 <sup>b</sup>	0.001 ± 0.00 <sup>b</sup>
2.5% KNO <sub>3</sub> +1 mM SA	0.72 ± 0.02 <sup>a</sup>	17.74 ± 2.09 <sup>a</sup>	0.001 ± 0.00 <sup>b</sup>
Levels of Significance	0.000	0.000	0.000

Means and standard error (SE) in same column having similar superscripts are statistically the same as shown by Duncan Multiple Range Test at P ≤ 0.05; SL stands for Seedling length; SB = Seedling biomass; RL = root length; MDA stands for malondialdehyde

## Discussions

Scarcity of water has been a crucial ecological burden that austere reduces crop harvest in arid and semi-arid biomes across the globe (Yan, 2015). Synergistic osmopriming and hormonal priming with 2.5% KNO<sub>3</sub>+3.5% SiO<sub>2</sub>, 3.5% SiO<sub>2</sub>+1 mM SA and 2.5% KNO<sub>3</sub>+1 mM SA improved germination attributes, seedling growth, antioxidant enzymes' activities as well as biochemical characteristics of FARO44 rice seedlings grown under PEG-induced water deficit stress. The findings of this study showed that osmo-hormonal priming improved germination index and decreased time taken for completion of germination (mean germination time) of FARO44 rice under PEG-induced drought stress. However, osmo-hormonal priming had no substantial effect in improving germination percentage of rice under PEG-induced drought. Enhancement of GI and decrease of MGT of rice was associated with

increased cell division, activation of important pre-germination processes in seeds, accelerated activities of antioxidants such as ascorbate peroxidase, superoxide dismutase and catalase as well as high content of proline, soluble sugar as well as improved imbibition of water in osmo-hormonal primed seeds. Heightened antioxidant activities in rice seedlings point out to resistance to water stress conditions. Osmotic potential regulation has been a strategy employed by plants to respond to prevalent water deficit circumstances (Farooq *et al.*, 2009). This study found that synergistic osmo-hormonal priming improved seedling growth of FARO44 rice under drought. Improved seedling growth might be due to the pivotal roles of osmo-hormonal priming in triggering cell division and elongation, restoration of impaired nucleic acid, production and galvanization of development proteins as well as heightened drought coping factors. Plants growing in drought threatened settings

*Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (Oryza sativa) FARO44 Under Water Deficit Conditions*



typically amass osmolytes like proline and soluble sugar that aid in improving osmotic modification (Yan, 2015). Previously, Farooq *et al.* (2013) found that ascorbic acid osmoprimed and hydroprimed wheat had improved biomass, seedling length plus phenolic and proline content.

It had been found that synergistic osmo-hormonal priming with 2.5% KNO<sub>3</sub>+3.5% SiO<sub>2</sub>, 3.5% SiO<sub>2</sub>+1 mM SA and 2.5% KNO<sub>3</sub>+1 mM SA improved seedling biomass under PEG-induced drought. Enhancement of biomass of rice seedling by osmo-hormonal priming was closely correlated to speedy germination initiated by hastened metabolic processes. Modulated hydrolases are paramount for production of metabolites that stimulate accelerated germination and uniform establishment of seedlings (Farooq *et al.*, 2006). Supporting these results, Farooq *et al.* (2013) found that ascorbic acid osmoprimed and hydroprimed wheat had improved biomass, seedling length. Equally, Rehman *et al.* (2011) stated that CaCl<sub>2</sub> osmoprimed rice had higher kernel mass as well as emergence index. Agreeing these results, PEG osmoprimed sorghum grown under limiting water supply was reported by Zhang *et al.* (2015) to have had improved dry biomass and vigour.

Synergistic osmo-hormonal priming with 2.5% KNO<sub>3</sub>+3.5% SiO<sub>2</sub>, 3.5% SiO<sub>2</sub>+1 mM SA and 2.5% KNO<sub>3</sub>+1 mM SA further improved activities of APX, CAT and SOD in rice seedlings grown under water constraint. While responding to stresses, plants deploy antioxidant systems consisting of catalase, glutathione, superoxide dismutase, peroxidase, superoxide dismutase and ascorbic acid to

convert the toxic ROS to harmless substances (Gill & Tuteja, 2010; Hussain *et al.*, 2016). Improved activities of antioxidants in rice seedlings suggested increased resistance to drought. Corroborating these results, the study by Yan (2015) found that KNO<sub>3</sub> soaked *Brassica rapa* grown under limited water had high POD, SOD and CAT activities.

It had been shown in this study that synergistic osmo-hormonal priming with 2.5% KNO<sub>3</sub>+3.5% SiO<sub>2</sub>, 3.5% SiO<sub>2</sub>+1 mM SA and 2.5% KNO<sub>3</sub>+1 mM SA enhanced proline content as well as lessened malondialdehyde content in rice seedlings subjected to limited supply of water. Proline is an essential osmolyte for osmotic adjustment in plants under stress, while malondialdehyde causes lipid peroxidation which is responsible for oxidative stress. Plants amass glycine betaine, proline, carbohydrate and soluble sugar as resistance tactics in limited water settings (Chowdhury *et al.*, 2017). Corroborating these results, Abdul Latef & Tran, (2016) stated that Si mediated maize grown in alkaline conditions had substantially higher total chlorophyll and soluble sugar. Salicylic acid and selenium osmo-hormonal primed rice cultivars under cold stress had high proline, glutathione, SOD, POD and CAT activities; low malondialdehyde content (Hussain *et al.*, 2016).

## Conclusions

Drought is a crucial ecological constraint that affects seed germination, growth of seedling and yield of cereals particularly rice across the globe. Synergistic osmo hormonal priming with 2.5% KNO<sub>3</sub>+3.5% SiO<sub>2</sub>, 3.5% SiO<sub>2</sub>+1 mM SA and 2.5%

*Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (Oryza sativa) FARO44 Under Water Deficit Conditions*

KNO<sub>3</sub>+1 mM SA proved effective in improving germination characteristics, growth of seedling as well as biochemical components of FARO44 rice subjected PEG-induced drought situations. Activities of radical destroying ascorbate peroxidase, superoxide dismutase and catalase were heightened in osmo-hormonal primed rice seedlings. Synergistic osmo-hormonal priming treatments also improved protein and proline contents in rice seedlings while decreased malondialdehyde content.

## References

- Abdul Latef, A. A., & Tran, L. P. (2016). Impacts of Priming with Silicon on the Growth and Tolerance of Maize Plants to Alkaline Stress. *Frontiers in Plant Science*, 7, 1–10. <https://doi.org/10.3389/fpls.2016.00243>
- Abdul-baki, A. A., & Anderson, J. D. (1970). Viability and Leaching of Sugars from Germinating Barley. *Crop Science*, 10, 3–6.
- Akinwale, M. G., Akinyele, B. O., Odiyi, A. C., Nwilene, F., Gregorio, G., & Oyetunji, O. E. (2012). Phenotypic Screening of Nigerian Rainfed Lowland Mega Rice Varieties for Submergence Tolerance. *Proceedings of World Congress on Engineering*, London, UK, 1, 4–9.
- Aloui, H., Souguir, M., & Hannachi, C. (2014). Determination of an optimal priming duration and concentration protocol for pepper seeds (*Capsicum annum L.*). *Acta Agriculturae Slovenica*, 103(2), 213–221. <https://doi.org/10.14720/aas.2014.103>
- Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (*Oryza sativa*) FARO44 Under Water Deficit Conditions
- .2.6
- Anosheh, H. P., Sadeghi, H., & Emam, Y. (2011). Chemical Priming with Urea and KNO<sub>3</sub> Enhances Maize Hybrids (*Zea mays L.*) Seed Viability under Abiotic Stress. *Journal of Crop Science and Biotechnology*, 14(4), 289–295.
- Ashraf, M., & Foolad, M. R. (2005). Pre-Sowing Seed Treatment-A Shotgun Approach to Improve Germination, Plant Growth, and Crop Yield Under Saline and Non-Saline Conditions. *Advances in Agronomy*, 88(05), 223–271. [https://doi.org/10.1016/S0065-2113\(05\)88006-X](https://doi.org/10.1016/S0065-2113(05)88006-X)
- Azeem, M., Iqbal, N., Kausar, S., Javed, M. T., Akram, M. S., & Sajid, M. A. (2015). Efficacy of silicon priming and fertigation to modulate seedling's vigor and ion homeostasis of wheat (*Triticum aestivum L.*) under saline environment. *Environmental Science and Pollution Research*, 22(18), 14367–14371. <https://doi.org/10.1007/s11356-015-4983-8>
- Bates, L.S., Waldren, R.P. & Teare, I.D. (1973). Rapid determination of free proline for water-stress studies. *Plant and Soil*, 39, 205–207.
- Bradford, M. M. (1976). A Rapid and Sensitive Method for the Quantitation Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Analytical Biochemistry*, 72, 248–254.
- Chowdhury, J., Karim, M., Khaliq, Q., & Ahmed, A. (2017). Effect of drought

- stress on bio-chemical change and cell membrane stability of soybean genotypes. *Bangladesh Journal of Agricultural Research*, 42(3), 475–485.  
<https://doi.org/10.3329/bjar.v42i3.34506>
- Chunthaburee, S., Sanitchon, J., Pattanagul, W., & Theerakulpisut, P. (2014). Alleviation of salt stress in seedlings of black glutinous rice by seed priming with spermidine and gibberellic acid. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, 42(2), 405–413.  
<https://doi.org/10.1583/nbha4229688>
- Dien, D. C., Mochizuki, T., & Yamakawa, T. (2019). Effect of various drought stresses and subsequent recovery on proline, total soluble sugar and starch metabolisms in Rice (*Oryza sativa* L.) varieties. *Plant Production Science*, 22(4), 530–545.  
<https://doi.org/10.1080/1343943X.2019.1647787>
- Farooq, M., Irfan, M., Aziz, T., Ahmad, I., & Cheema, S. A. (2013). Seed Priming with Ascorbic Acid Improves Drought Resistance of Wheat. *Journal of Agronomy and Crop Science*, 199(1), 12–22.  
<https://doi.org/10.1111/j.1439-037X.2012.00521.x>
- Farooq, M., Wahid, A., Kobayashi, N., Fujita, D., & Basra, S. M. A. (2009). Plant drought stress: Effects, mechanisms and management. *Sustainable Agriculture*, 29, 153–188.  
[https://doi.org/10.1007/978-90-481-2666-8\\_12](https://doi.org/10.1007/978-90-481-2666-8_12)
- Farooq, Muhammad, Basra, S. M. A., Tabassum, R., & Afzal, I. (2006). Enhancing the Performance of Direct Seeded Fine Rice by Seed Priming. *Plant Production Science*, 9(4), 446–456.  
<https://doi.org/10.1626/pp.s.9.446>
- Farooq, M., Basra, S. M. A., Wahid, A., & Ahmad, N. (2010). Changes in Nutrient-Homeostasis and Reserves Metabolism During Rice Seed Priming: Consequences for Seedling Emergence and Growth. *Agricultural Sciences in China*, 9(2), 191–198.  
[https://doi.org/10.1016/S1671-2927\(09\)60083-3](https://doi.org/10.1016/S1671-2927(09)60083-3)
- Ghobadi, M., Abnavi, M. S., Honarmand, S. J., & Ghobadi, M. E. (2012). Effect of Hormonal Priming (GA<sub>3</sub>) and Osmopriming on Behavior of Seed Germination in Wheat (*Triticum aestivum* L.). *Journal of Agricultural Science*, 4(9), 244–250.  
<https://doi.org/10.5539/jas.v4n9p244>
- Gill, S. S., & Tuteja, N. (2010). Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. *Plant Physiology and Biochemistry*, 48(12), 909–930.  
<https://doi.org/10.1016/j.plaphy.2010.08.016>
- Goswami, A., Banerjee, R., & Raha, S. (2013). Drought resistance in rice seedlings conferred by seed priming: Role of the antioxidant defense mechanisms. *Protoplasma*, 250(5), 1115–1129.  
<https://doi.org/10.1007/s00709-013-0487-x>
- Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (Oryza sativa) FARO44 Under Water Deficit Conditions*



- Heath, R. L., & Packer, L. (1968) Photoperoxidation in isolated Chloroplasts I. Kinetics and Stoichiometry of Fatty Acid Peroxidation. *Archives of Biochemistry & Biophysics*, 125, 189–198.
- Hussain, S., Khan, F., Hussain, H. A., & Nie, L. (2016). Physiological and Biochemical Mechanisms of Seed Priming-Induced Chilling Tolerance in Rice Cultivars. *Frontiers in Plant Science*, 7, 1–14. <https://doi.org/10.3389/fpls.2016.00116>
- Hussain, S., Zheng, M., Khan, F., Khaliq, A., Fahad, S., Peng, S., Huang, J., Cui, K., & Nie, L. (2015). Benefits of rice seed priming are offset permanently by prolonged storage and the storage conditions. *Scientific Reports*, 5, 8101. <https://doi.org/10.1038/srep08101>
- IPCC, 2007: Fourth assessment report: synthesis. [http://www.ipcc.ch/pdf/assessment-report/ar4/syr/ar4\\_syr.pdf](http://www.ipcc.ch/pdf/assessment-report/ar4/syr/ar4_syr.pdf)
- Ji, K., Wang, Y., Sun, W., Lou, Q., Mei, H., Shen, S., & Chen, H. (2012). Drought-responsive mechanisms in rice genotypes with contrasting drought tolerance during reproductive stage. *Journal of Plant Physiology*, 169(4), 336–344. <https://doi.org/10.1016/j.jplph.2011.10.010>
- Jisha, K. C., & Puthur, J. T. (2016). Seed Priming with Beta-Amino Butyric Acid Improves Abiotic Stress Tolerance in Rice Seedlings. *Rice Science*, 23(5), 242–254. <https://doi.org/10.1016/j.rsci.2016.08.002>
- Jisha, K., Vijayakumari, K. C., & Puthur, J. T. (2013). Seed priming for abiotic stress tolerance: an overview. *Acta Physiol Plant*, 35, 1381–1396. <https://doi.org/10.1007/s11738-012-1186-5>
- Kamoshita, A., Rodriguez, R., Yamauchi, A., & Wade, L. (2004). Genotypic Variation in Response of Rainfed Lowland Rice to Prolonged Drought and Rewatering. *Plant Production Science*, 7(4), 406–420. <https://doi.org/10.1626/pp.s.7.406>
- Kaufmann, M. R. (1973). The Osmotic Potential of Polyethylene Glycol 60001. *Plant Physiology*, 51, 914–916.
- Kaya, M. D., Okcu, G., Atak, M., Cikili, Y., & Kolsarıcı, O. (2006). Seed treatments to overcome salt and drought stress during germination in sunflower (*Helianthus annuus* L.). *European Journal of Agronomy*, 24, 291–295. <https://doi.org/10.1016/j.eja.2005.08.001>
- Khan, M. N., Zhang, J., Luo, T., Liu, J., Rizwan, M., Fahad, S.,...Hu, L. (2019). Seed priming with melatonin coping drought stress in rapeseed by regulating reactive oxygen species detoxification: Antioxidant defense system, osmotic adjustment, stomatal traits and chloroplast ultrastructure perseveration. *Industrial Crops &*

*Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (Oryza sativa) FARO44 Under Water Deficit Conditions*

- Products*, 140, 111597.  
<https://doi.org/10.1016/j.indcrop.2019.111597>
- Liu, H., Hussain, S., Zheng, M., Peng, S., Zheng, M., Peng, S., & Huang, J. (2016). Dry direct-seeded rice as an alternative to transplanted-flooded rice in Central China. *Agronomy for Sustainable Development*, 35(1), 285–294.  
<https://doi.org/10.1007/s13593-014-0239-0>
- Muchate, N. S., Nilima, S. R., Suprasanna, P., & Nikam, T. D. (2019). NaCl induced salt adaptive changes and enhanced accumulation of 20-hydroxyecdysone in the in vitro shoot cultures of *Spinacia oleracea* (L.). *Scientific Reports*, 9, 1–10.  
<https://doi.org/10.1038/s41598-019-48737-6>
- Nakano, Y., & Asada, K. (1981). Hydrogen Peroxide is Scavenged by Ascorbate-specific Peroxidase in Spinach Chloroplasts. *Plant & Cell Physiology*, 22(5), 867–880.
- Oluwaseyi, A. B., Nehemiah, D., & Zuluqurineen, S. B. (2016). Genetic Improvement of Rice in Nigeria for Enhanced Yield and Grain Quality - A Review. *Asian Research Journal of Agriculture*, 1(3), 1–18.  
<https://doi.org/10.9734/ARJA/2016/28675>
- Pandey, V., & Shukla, A. (2015). Acclimation and Tolerance Strategies of Rice under Drought Stress. *Rice Science*, 22(4), 147–161.  
<https://doi.org/10.1016/j.rsci.2015.04.001>
- Potassium nitrate, Silicon dioxide and Salicylic Acid Mediation Increase Seedling Growth, Biochemical Characteristics and Antioxidant Enzymes Activities in Rice (Oryza sativa) FARO44 Under Water Deficit Conditions*
- Paparella, S., Araújo, S. S., Rossi, G., Wijayasinghe, M., Carbonera, D., & Balestrazzi, A. (2015). Seed priming: state of the art and new perspectives. *Plant Cell Reports*, 34(8), 1281–1293. <https://doi.org/10.1007/s00299-015-1784-y>
- Price, A. H., Atherton, N. M., & Hendry, G. A. F. (1989). Plants under Drought-Stress Generate Activated Oxygen. *Free Radical Resources Communication*, 8(1), 61–66.
- Quan, N. T., Anh, L. H., Khang, D. T., & Tuyen, P. T. (2016). Involvement of Secondary Metabolites in Response to Drought Stress of Rice (*Oryza sativa* L.). *Agriculture*, 6(23), 2–14.  
<https://doi.org/10.3390/agriculture6020023>
- Refli, & Purwestri, Y. A. (2016). The response of antioxidant genes in rice (*Oryza sativa* L.) seedling Cv. Cempo Ireng under drought and salinity stresses. *AIP Conference Proceedings*, 1744.  
<https://doi.org/10.1063/1.4953521>
- Rehman, H. U., Maqsood, S., Basra, A., & Farooq, M. (2011). Field appraisal of seed priming to improve the growth, yield, and quality of direct seeded rice. *Turkish Journal of Agriculture & Forestry*, 35, 357–365.  
<https://doi.org/10.3906/tar-1004-954>
- Shehab, G. G., Ahmed, O. K., & El-Beltagi, H. S. (2010). Effects of various chemical agents for alleviation of drought stress in rice plants (*Oryza sativa* L.). *Notulae*

- Botanicae Horti Agrobotanici Cluj-Napoca*, 38(1), 139–148.  
<https://doi.org/10.15835/NBHA3813627>
- Thomas, S., Anand, A., & Chinnusamy, V. (2013). Magnetopriming circumvents the effect of salinity stress on germination in chickpea seeds. *Acta Physiol Plant*, 35, 3401–3411.  
<https://doi.org/10.1007/s11738-013-1375-x>
- Yan, M. (2015). Seed priming stimulates germination and early seedling growth of Chinese cabbage under drought stress. *South African Journal of Botany*, 99, 88–92.  
<https://doi.org/10.1016/j.sajb.2015.03.195>
- Zhang, F., Yu, J., Johnston, C. R., Wang, Y., Zhu, K., & Lu, F. (2015). Seed Priming with Polyethylene Glycol Induces Physiological Changes in Sorghum (*Sorghum bicolor* L. Moench) Seedlings under Suboptimal Soil Moisture Environments. *PLoS ONE*, 10(10), 1–15.  
<https://doi.org/10.1371/journal.pone.0140620>
- Zheng, M., Tao, Y., Hussain, S., Jiang, Q., & Peng, S. (2016). Seed priming in dry direct-seeded rice: consequences for emergence, seedling growth and associated metabolic events under drought stress. *Plant Growth Regulation*, 78(2), 167–178.  
<https://doi.org/10.1007/s10725-015-0083-5>